

Distribution and antibiogram pattern of *Acinetobacter* infections in Shahid Mohammadi Hospital, Bandar Abbas, Iran

Pyrooz Poorzargar¹ Sedigheh Javadpour² Afsaneh Karmostaji²

Infectious and Tropical Diseases Research Center¹, Hormozgan University of Medical Sciences, Bandar Abbas, Iran. Deptment of Microbiology², Infectious and Tropical Diseases Research Center, Hormozgan Health Istiute, Hormozgan University of Medical Sciences, Bandar Abbas, Iran.

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Original Article

Abstract

Introduction: *Acinetobacter* species are important opportunistic pathogens, widely spread in hospitals' environment and responsible for different health care associated infections. Because of its ability to rapidly develop resistance to the major groups of antibiotics, treatment of *Acinetobacter* infections is difficult and antibiotic susceptibility tests can help in choosing the best antibiotics, decreasing the cost and duration of hospitalization. The goals of this study were to determine frequency and antimicrobial susceptibility pattern of *Acinetobacter* species, clinical parameters and outcomes of patients, in Shahid Mohammadi hospital, Bandar Abbas.

Methods: Between April 2010 and March 2011, a total of 2132 positive cultures were obtained from various clinical specimens of hospitalized patients. Suspicious isolates of *Acinetobacter* were identified by routine microbiological methods. Antibiogram patterns of isolates for 12 currently used antibiotics were determined by Kirby-Bauer method. Clinical and microbiological data of patients was analyzed by SPSS 16 software.

Results: A total of 68 (3.2%) *Acinetobacter* species was isolated. *Acinetobacter* isolates was mostly obtained from ICU (24 cases, 35.8%) and emergency (12 cases, 17.9%) wards, and trachea was the major site of infection (41.2%). Colistin with 83.7% susceptibility rate was the most effective antibiotic, followed by ofloxacin 47.4% and chloramphenicol 39.5%. A high rate of resistance was observed to meropenem (98.1%), and cefepime (90.4%). Mortality rate was 14.7% in patients, mostly because of bacteremia.

Conclusion: Because of its serious infections and high-drug resistance, continuous monitoring of antimicrobial susceptibility and strict adherence to infection guidelines are essential to prevent and decrease *Acinetobacter* infections.

Key words: *Acinetobacter*, Microbiasensitivity Test, Intensive Care Units

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Introduction:

In a recent report of the Infectious Diseases Society of America, three categories of MDR gram-negative bacilli, namely, extended-spectrum

cephalosporin-resistant *Escherichia coli* and *Klebsiella spp.*, MDR *Pseudomonas aeruginosa*, and carbapenem-resistant *Acinetobacter spp.* are the main concerns of antibiotic therapy (1).

Acinetobacter species are increasingly important nosocomial Pathogen (2). Before the 1970s, *Acinetobacter* were mostly isolated from postsurgical urinary tract infections. The significant improvement in cardiovascular recovery and the use of invasive techniques or artificial devices during the last 30 years has changed the types of infection caused by these bacteria. Since the 1980s, *Acinetobacter* species has spread rapidly among patients in intensive care units (3).

Today, *Acinetobacter* species account for 9% of nosocomial infections (4). *A. baumannii* is intrinsically resistant to some beta-lactam antibiotics and has ability to acquire resistance to all commercially available antimicrobial agents. In fact, carbapenem-resistant *A. baumannii* has been identified as one of the six pathogens *Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella* species, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Enterobacter* (ESKAPE) species responsible for an increasing number of nosocomial infections in the United States (5).

The six bad bugs known as ESKAPE bacteria are among the biggest threats infectious diseases physicians face today. *Acinetobacter* species are excellent biofilm producing bacteria, which facilitate their survival in hospital environments and are frequently found on the skin and in the respiratory and urinary tracts of hospitalized patients (6). Infected or colonized patients are the main reservoir of these bacteria. Because of reduced acid secretion, bacterial overgrowth in the stomach of ICU patients can lead to the development of nosocomial infections (4). The main route of transmission is the hands of hospital care workers and inanimate objects act as an intermediate reservoir between the hands of hospital workers and the patients (7). Study on 3 Army hospitals in Iran, showed that *A. baumannii* is responsible of 9% of infections in ICU patients (8).

An Iranian review study, imply an increasing in antibiotic-resistant *A. baumannii* strains from 2001 to 2013. The prevalence of MDR strains also have been increased from 50% in 2001-2007 to 74% in 2010-2011, with a mean prevalence of 71.2% (9).

The severity of *Acinetobacter* infections depend upon the site of infection and the patient's susceptibility as a result of underlying disease (10).

The purpose of this study is to provide data on the frequency, outcome, and antibiotic susceptibility of *Acinetobacter* infections, with the goal of improving its management.

Methods:

This retrospective, descriptive cross-sectional study was carried out between April 2010 and March 2011, on 2132 different clinical specimens, submitted to Shahid Mohammadi Hospital (a 400-bed hospital serving of over two million people). Laboratory isolation of the bacteria was carried out by culturing the specimens on appropriate bacteriological media, including Blood agar, Chocolate agar, thioglycollate, EMB or McConkey agar. Cultures were incubated at 37°C for 24 - 48 h. Blood samples were inoculated in Trypticase soy broth bottles and incubated for at least 7 days at 37°C. Identification of *Acinetobacter* isolates was performed by routine microbiological methods using Gram staining, oxidase, TSI and motility results (11).

In vitro antibiotic susceptibility of the isolates to 12 antibiotics was determined by the Kirby-Bauer disc diffusion method, on Mueller-Hinton agar. Antibiogram discs contained the following antibiotics, at the specific concentrations indicated in parentheses: Imipenem (10µg), meropenem (10µg), colistin (25µg) ceftriaxone (30µg), cefepime (30µg), ciprofloxacin (5 µg), ceftazidime (30µg), cefazolin (30µg), ofloxacin (5µg), amikacin (30µg), gentamicin (10µg), cotrimoxazol (25µg) which were purchased from PadTanTeb, Iran. According to the guidelines of Clinical and Laboratory Standards Institute (CLSI), cell suspension inoculates were prepared from 18 - 24 h fresh, pure cultures, in 0.85% sterile saline and adjusted to match a 0.5 McFarland standard tubes. The criteria proposed by CLSI were applied for interpreting the results (12).

Leukocyte count was done by automated leukocyte analyzer method, C-reactive protein (CRP) by qualitative method of latex-CRP (ENiSon, ENiSon Lab, Tehran, Iran), and Erythrocyte sedimentation rate (ESR) by Westergren method. Results of latex-CRP tests were reported according to the presence or absence of agglutination and size of agglutinated droplets on

microscopic examination: no agglutination was considered negative; small-sized agglutinated droplets, 1+; medium-sized agglutinated droplets, 2+; and large-sized agglutinated droplets, 3+.

Other variables included in the study were age, sex, dates of admission and discharge, type of clinical specimen from which the *Acinetobacter* strains were isolated and death during hospitalization. Analysis of data was performed by SPSS 16. Significance was defined as $P \leq 0.05$.

Results:

Out of 2132 positive cultures, a total of 68 (3.18%) *Acinetobacter* species was isolated from various clinical specimens. 49 (72.1) of them were isolated from men and 19 (27.9%) obtained from women.

Trachea was the major site of infection (41.8%), followed by blood stream and urinary tract (16.18%) distinctly and wounds swabs (13.2%). Table 1 shows the occurrence rates of *Acinetobacter* isolates by site of infection.

Table 1. Frequency of *Acinetobacter* isolates by body sites

| Sample | Frequency | Percent |
|-----------------|-----------|------------|
| Tracheal tube | 28 | 41.18 |
| Blood | 11 | 16.18 |
| Urine | 11 | 16.18 |
| Wound | 9 | 13.24 |
| Sputum | 4 | 5.89 |
| CSF | 1 | 1.47 |
| Ear discharge | 1 | 1.47 |
| Catheter | 1 | 1.47 |
| Drain discharge | 1 | 1.47 |
| Eye discharge | 1 | 1.47 |
| Total | 68 | 100 |

Table 2. Distribution of *Acinetobacter* species according to different wards of hospital

| Wards | Frequency | Percentage |
|--------------------|-----------|------------|
| General ICU | 24 | 35.30 |
| Internal Emergency | 12 | 17.65 |
| Internal 1 | 10 | 14.71 |
| Surgery 1&2 | 5 | 7.36 |
| Internal 2 | 4 | 5.89 |
| Neurosurgery ICU | 4 | 5.89 |
| Internal 3 | 3 | 4.42 |
| Orthopaedic | 3 | 4.42 |
| Cardio Surgery | 2 | 2.95 |
| Burn | 1 | 1.47 |
| Total | 68 | 100 |

Acinetobacter isolates were mostly obtained from ICU (24 cases, 35.8%), followed by Internal emergency (12 cases, 17.9%) and internal 1 (10 cases, 14.7%) wards patients. The distribution of *Acinetobacter* species according to different wards of hospital is presented in Table 2.

Table 3 indicates antibiogram pattern of *Acinetobacter* species to tested antibiotics. As it is considered a high rate of resistance was observed to cefepime (90.4%), ceftazidime (89.4%), ceftriaxone (80.3%), cefazolin (75%) and amikacin (70.8%). Colistin with 83.7% susceptibility rate was the most effective antibiotic.

The data of peripheral leukocyte count (WBC), serum C-reactive protein (CRP) level and erythrocyte sedimentation rate (ESR) of the patients was extracted from their medical records, (not found for all patients) and is shown in Table 4.

The Median time of hospitalization in 88.2% of patients were more than 7 days. About 78.4% of infected patients were febrile. Ten deaths were reported in the infected patients and mortality rate was 14.7%. Septicemia was the major cause of death (5 cases). Other causes of decease were respiratory failure (2 cases) and electric shock, brain injury and pulmonary emboli (Table 5).

Table 3. Antibigram pattern of *Acinetobacter* species (expressed as percentage)

| Resistant | Intermediate | Sensitive | Number | Antibiotic |
|---------------|--------------|-----------|--------|------------|
| Ciprofloxacin | 59 | 37.29 | 1.70 | 61.02 |
| Imipenem | 55 | 32.73 | 3.64 | 63.64 |
| Meropenem | 52 | 1.92 | 0 | 98.08 |
| Amikacin | 65 | 21.16 | 3.08 | 70.77 |
| Gentamicin | 64 | 35.94 | 6.25 | 57.82 |
| Ceftazidime | 47 | 8.41 | 2.13 | 89.37 |
| Ceftriaxone | 66 | 18.19 | 1.52 | 80.31 |
| Cotrimoxazole | 54 | 35.19 | 1.86 | 62.97 |
| Colistin | 49 | 83.67 | 0 | 16.32 |
| Cefazolin | 23 | 21.74 | 0 | 78.26 |
| Cefepime | 52 | 9.62 | 0 | 90.38 |
| Ofloxacin | 19 | 47.37 | 5.26 | 47.37 |

Table 4. Infection and inflammation-related hematological parameters

| | CRP | WBC | | ESR | |
|----------|-----------|-------------|------------|-------|----------|
| | No. | No. | No. | No. | No. |
| Negative | 6 (23.1%) | > 5000 | 0 | ≥20 | 3 (12%) |
| +1 | 7 (26.9%) | 5000-11000 | 15 (29.5%) | 21-30 | 2 (8%) |
| +2 | 8 (30.8%) | 11000-20000 | 25 (49%) | <30 | 20 (80%) |
| +3 | 5 (19.2%) | < 20000 | 11 (21.5%) | | |

Table 5. Deaths according to the final diagnosis

| Ward | Body temperature (°C) | Final diagnosis | Sex | Age |
|--------------------------|-----------------------|--------------------------------|--------|-----|
| 1 ICU-General | 38.5 | MT, Pul emboli | Male | 28 |
| 2 ICU-G | 38 | MT, DAI, TE fistula | Male | 18 |
| 3 ICU-G | 39 | MT, Sepsis | Male | 23 |
| 4 Cardiothoracic Surgery | 38.2 | CAD, Sepsis | Female | 58 |
| 5 ICU-G | 38.5 | Respiratory failure, ARDS | Female | 29 |
| 6 ICU-G | 39 | Electrical injury | Male | 23 |
| 7 ICU-G | 38 | MT, DAI | Male | 12 |
| 8 Cardiothoracic Surgery | 37.8 | COPD, CAD, Respiratory failure | Female | 81 |
| 9 Internal ward | 38.6 | MT, Sepsis | Male | 55 |
| 10 ICU-G | 38 | TF, Sepsis, DIC | Male | 19 |

MT: Multiple trauma DAI: Diffuse axonal injury

ARDS: Acute respiratory distress syndrome

DIC: Disseminated intravascular coagulation

TE fistula: Tracheoesophageal fistula

COPD: Chronic obstructive pulmonary disease

ICU-G: ICU general

CAD: Coronary artery disease

TF: Tetralogy fallot

Conclusion:

In the present study *Acinetobacter* species accounted for 3.2% of identified microorganisms, in clinical cultures, which is lower than other reports from Iran, in which 23.5% and 16.1% of isolates were found to be *Acinetobacter* species in 1993 (13) and in 2004 in Tehran (14).

In Canadian hospitals, *Acinetobacter* made 0.7% of all isolates in intensive care unit in 2005-

2006 (15). This discrepancy may be due to the different detection methods and standards of hygiene in the hospitals.

According to our results trachea was the major site of infection (41.2%). In a study on multidrug *A. baumannii* infection during an hospital outbreak, respiratory isolates were recovered from 53% of the patients (16). In Dash study maximum (56.9%) isolates of *A. baumannii* were obtained from pus

swabs of elderly age inpatients. longer duration of stay in the hospital, associated co-morbidity, and invasive procedure were found to be significant risk factors in the setup investigated. Comparable to their findings, blood (13.1%) and urine (12.4%), we detected that 16.2% of isolates were attributed to blood and urine cultures, distinctly (17).

High sensitivity of our isolates to colistin is in consistent with other studies in Iran and other countries around the world (18-22). In Amudhan study in India all of the *A. baumannii* isolates were resistant to imipenem and meropenem (23). Many studies have indicated increasing carbapenem-resistant *A. baumannii*, from 8% in 2003 to 52% and 74% in 2005 and 96% in 2007(24-25). In Iran it was reported between 49.2% to 52.5% during 2008-2011 (26-27).

Assessment of CRP, ESR and WBC are reliable methods to diagnose bacterial inflammations, CRP is an innate immune system response to acute and chronic infections and ESR gives a nonspecific measure of inflammation. The WBC total count includes information on five types that can indicate the type of infection: eosinophils, neutrophils, leukocytes, basophils, and monocytes (28).

In the present study, ESR above 30 was detected in 80% of the patients and 76.9% of patients had CRP 1+. The total count of leukocytes was also more than 11000 in 70.5% of patients, as an indication of bacterial inflammation. As it is established, one of the important quantitative factors in nosocomial infections is duration of hospitalization. Respectively, in our study 45 patients (88.2%) were hospitalized for more than 7 days.

In a study on a hospital outbreak by multi-drug resistant *A. baumannii*, 43.3% of mortalities was because of respiratory infections and the significant quantitative variable was the number of days of hospitalization (22). In another study, Pinyo and co-workers indicated that infection or colonization by *A. baumannii* was associated with an additional hospital stay of 14 days with mortality rate of 24.3% (29).

About 35.8% of isolates in our study were from ICU patients, signifying that ICU patients are at greatest risk of infection. Correspondingly a survey by the Health Protection Agency in England found that 54% of patients with *Acinetobacter* bacteraemia

were hospitalized in ICUs between 1976 and 1990 (4).

In our study, mortality rate was 14.7% and septicemia was the major cause of death. Of course it is difficult to prove that all mortalities was due to *Acinetobacter* infection. In another study on clinical outcomes of patients with bloodstream infection caused by carbapenem-resistant *A. baumannii*, mortality rate was 41%. Risk factors associated with mortality were intensive care unit stay, malignancy, and presence of fever and/or hypotension (30). Data from other studies suggests that the crude or related mortality rate ranges from 20% to 60% (1). In one review paper, incidence of nosocomial bacteremia in ICU patients was reported from 1% to as high as 6.5%, compare to 0.65% in other hospitalized patients, and the mortality rate from *Acinetobacter* meningitis was between 15% to 71% (31).

Del Mar Toma study indicated that the mortality among patients with *A. baumannii* bacteremia was 19.6% (16). Munoz and co-workers founded that presence of fever and/or hypotension were associated with a higher death rate (30). These data confirms that a high long hospitalization is attributable to *Acinetobacter* infections and increase of mortality rate.

We believe that our study provides reliable information on the frequency, outcome, and antibiotic resistance pattern of *Acinetobacter* species. Our study was limited by the lack of severity scores (ie, mechanical ventilation, tracheostomy, immunosuppressions associated with transplantation, and malignancies).

As a final point, due to the important of *Acinetobacter* infections and antimicrobial resistance, the tracing of the infectious sources is of great significance for the control and prevention managements.

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References:

- Giske CG, Monnet DL, Cars O, Carmeli Y, ReAct-Action on Antibiotic. Clinical and economic impact of common multidrug-resistant gram-negative bacilli. *Antimicrobial Agents and Chemotherapy*. 2008;52(3):813-821.
- Cisneros JM, Rodriguez-Bano J. Nosocomial bacteremia due to *Acinetobacter baumannii*: epidemiology, clinical features and treatment. *Clinical microbiology and infection: the official publication of the European Society of Clinical Microbiology and Infectious Diseases*. 2002;8(11):687-693.
- Joly-Guillou ML. Clinical impact and pathogenicity of *Acinetobacter*. *Clinical Microbiology and Infection*. 2005;11(11):868-873.
- Joly-Guillou ML. Clinical impact and pathogenicity of *Acinetobacter*. *Clinical microbiology and infection: the official publication of the European Society of Clinical Microbiology and Infectious Diseases*. 2005;11(11):868-73.
- Shields RK, Clancy CJ, Gillis LM, Kwak EJ, Silveira FP, Massih RC, et al. Epidemiology, clinical characteristics and outcomes of extensively drug-resistant *Acinetobacter baumannii* infections among solid organ transplant recipients. *PLoS One*. 2012;7(12):e52349.
- Kim UJ, Kim HK, An JH, Cho SK, Park KH, Jang HC. Update on the Epidemiology, Treatment, and Outcomes of Carbapenem-resistant *Acinetobacter* infections. *Chonnam Med J*. 2014;50(2):37-44.
- del Mar Tomas M, Cartelle M, Pertega S, Beceiro A, Llinares P, Canle D, et al. Hospital outbreak caused by a carbapenem-resistant strain of *Acinetobacter baumannii*: patient prognosis and risk-factors for colonisation and infection. *Clin Microbiol Infect*. 2005;11(7):540-546.
- Mohammadi-Mehr M, Feizabadi M. Antimicrobial resistance pattern of Gram-negative bacilli isolated from patients at ICUs of Army hospitals in Iran. *Iranian Journal of Microbiology*. 2011;3(1):26-30.
- Moradi J, Hashemi FB, Bahador A. Antibiotic resistance of *Acinetobacter baumannii* in Iran: a systemic review of the published literature. *Osong Public Health and Research Perspectives*. 2015;6(2):79-86.
- Bayramoglu G, Kaya S, Besli Y, Cakir E, Can G, Akineden D, et al. Molecular epidemiology and the clinical significance of *Acinetobacter baumannii* complex isolated from cerebrospinal fluid in neurosurgical intensive care unit patients. *Infection*. 2012;40(2):163-172.
- Winn WJ, Allen S, Janda W, Koneman E, Procop G, Schreckenberger P, et al. 6th ed. Philadelphia. Lippincott Williams & Wilkins; 2006.
- Performance standards for antimicrobial susceptibility testing; Twenty-first informational supplement. M100-S21. 2011;31(1).
- Nouroozi J ha, Mohammadi M. *Acinetobacter* as a nosocomial pathogen. *Iranian Journal of Infectious and Tropical Medicine (IJITM)*. 2004;24(9):1-5.
- Hadadi A, Rasoulinejad M, Zia basharhagh N. Investigation of the frequency of gram negative nosocomial infections and evaluation of antimicrobial resistance patterns by E-Test and Disk diffusion methods in Sina hospital. 2008;13(1):51-57.
- Zhanel G, Decorby M, Laing N, Weshnoweski B, Vashisht R, Tailor F, et al. Antimicrobial Resistant Pathogens in Intensive Care Units in Canada: Results of the Canadian National Intensive Care Unit (CAN-ICU) Study 2005-2006. *Journal of Antimicrobial agents and chemotherapy*. 2008;52(4):1430-1437.
- del Mar Tomas M, Cartelle M, Pertega S, Beceiro A, Llinares P, Canle D, et al. Hospital outbreak caused by a carbapenem-resistant strain of *Acinetobacter baumannii*: patient prognosis and risk-factors for colonisation and infection. *Clinical microbiology and infection: the official publication of the European Society of Clinical Microbiology and Infectious Diseases*. 2005;11(7):540-546.
- Dash M, Padhi S, Pattnaik S, Mohanty I, Misra P. Frequency, risk factors, and antibiogram of *Acinetobacter* species isolated from various clinical samples in a tertiary care hospital in Odisha, India. *Avicenna Journal of Medicine*. 2013;3(4):97-102.
- Dizbay M, Altuncekic A, Sezer BE, Ozdemir K, Arman D. Colistin and tigecycline susceptibility among multidrug-resistant

- Acinetobacter baumannii isolated from ventilator-associated pneumonia. *Int J Antimicrob Agents* 2008;32(1):29-32.
19. Karmostaj A, Najarpeerayeh S, Salmanian AH. Emergence of Tigecycline Resistant Acinetobacter baumannii From an Intensive Care Unit (ICU) in Tehran. *Jundishapur J Microbiol.* 2013;6(3):215-219.
 20. Feizabadi MM, Fathollahzadeh B, Taherikalani M, Rasoolinejad M, Sadeghifard N, Aligholi M, et al. Antimicrobial susceptibility patterns and distribution of blaOXA genes among Acinetobacter spp. Isolated from patients at Tehran hospitals. *Japanese Journal of Infectious Diseases.* 2008;61(4):274-278.
 21. Smolyakov R, Borer A, Riesenber K, Schlaeffer F, Alkan M, Porath A, et al. Nosocomial multi-drug resistant Acinetobacter baumannii bloodstream infection: risk factors and outcome with ampicillin- sulbactam treatment. *J Hospital Infection.* 2003;54(1):32-38.
 22. Shutt CK, Pounder JI, Page SR, Schaecher BJ, Woods GL. Clinical evaluation of the DiversiLab microbial typing system using repetitive-sequence-based PCR for characterization of Staphylococcus aureus strains. *Journal of Clinical Microbiology.* 2005;43(3):1187-1192.
 23. Amudhan SM, Sekar U, Arunagiri K, Sekar B. OXA beta-lactamase-mediated carbapenem resistance in Acinetobacter baumannii. *Indian Journal of Medical Microbiology.* 2011;29(3):269-274.
 24. Qi C, Malezynski M, Parker M, Scheetz MH. Characterization of genetic diversity of carbapenem-resistant Acinetobacter baumannii clinical strains collected from 2004 to 2007. *J Clin Microbiol.* 2008;46(3):1106-1109.
 25. Stoeva T, Higgins PG, Savov E, Markovska R, Mitov I, Seifert H. Nosocomial spread of OXA-23 and OXA-58 beta-lactamase-producing Acinetobacter baumannii in a Bulgarian hospital. *J Antimicrob Chemother.* 2009;63(3):618-620.
 26. Shahcheraghi F, Abbasalipour M, Feizabadi M, Ebrahimipour G, Akbari N. Isolation and genetic characterization of metallo-beta-lactamase and carbapenemase producing strains of Acinetobacter baumannii from patients at Tehran hospitals. *Iran J Microbiol.* 2011;3(2):68-74.
 27. Taherikalani M, Fathollahzadeh B, Emaneini M, Soroush S, Feizabadi, MM. Distribution of different carbapenem resistant clones of Acinetobacter baumannii in Tehran hospitals. *New Microbiol.* 2009;32(3):265-271.
 28. Wang L, Yang B, Yin B, Zhang Z, Zhang L, Tang L, et al. Clinical significance of PCT, CRP, ESR, WBC count as predictors in postoperative early infectious complications with fever after posterior lumbar internal fixation]. *Zhongguo gu shang. China Journal of Orthopaedics and Traumatology.* 2015;28(1):66-70.
 29. Rattanaumpawan P, Ussavasodhi P, Kiratisin P, Aswapokee N. Epidemiology of bacteremia caused by uncommon non-fermentative gram-negative bacteria. *BMC Infectious Diseases.* 2013;13:167.
 30. Munoz-Price LS, Zembower T, Penugonda S, Schreckenberger P, Lavin MA, Welbel S, et al. Clinical outcomes of carbapenem-resistant Acinetobacter baumannii bloodstream infections: study of a 2-state monoclonal outbreak. *Infection Control and Hospital Epidemiology.* 2010;31(10):1057-1062.
 31. Sligl W, Taylor G, Brindley PG. Five years of nosocomial Gram-negative bacteremia in a general intensive care unit: epidemiology, antimicrobial susceptibility patterns, and outcomes. *International Journal of Infectious Diseases.* 2006;10(4):320-325.

پراکندگی و الگوی حساسیت آنتی‌بیوتیکی عفونت‌های آسیتوباکتر در بیمارستان شهید محمدی بندرعباس

پیروز پورزرگر^۱ صدیقه جوادیپور^۲ افسانه کرمستجی^۲

^۱ پزشک عمومی، مرکز تحقیقات بیماریهای عفونی و گرمسیری، پژوهشکده سلامت هرمزگان، دانشگاه علوم پزشکی هرمزگان، بندرعباس، ایران. ^۲ گروه میکروبیولوژی، مرکز تحقیقات بیماریهای عفونی و گرمسیری، پژوهشکده سلامت هرمزگان، دانشگاه علوم پزشکی هرمزگان، بندرعباس، ایران.

مجله پزشکی هرمزگان سال بیستم شماره ششم ۹۵ صفحات ۴۰۳-۳۹۶

چکیده

مقدمه: آسیتوباکترها، باکتریهای فرصت طلب مهمی هستند که به طور وسیعی در بیمارستانها پراکنده و عامل عفونتهای بیمارستانی می‌باشند و چون به سرعت به گروه‌های اصلی آنتی‌بیوتیک‌ها مقاوم می‌شوند، درمان آنها مشکل است. الگوی حساسیت آنتی‌بیوتیکی می‌تواند در انتخاب آنتی‌بیوتیک‌ها، موثر و هزینه درمان و مدت بستری را کاهش دهد. هدف این مطالعه، تعیین فراوانی و الگوی حساسیت آنتی‌بیوتیکی گونه‌های آسیتوباکتر در بیمارستان شهید محمدی بندرعباس بود.

روش کار: در این مطالعه توصیفی مقطعی گذشته نگر، تعداد ۲۱۳۲ کشت مثبت حاصل از نمونه‌های مختلف بالینی بیماران بستری بررسی و کلنی‌های آسیتوباکتر با روش‌های میکروبی متداول شناسایی شدند. الگوی مقاومت آنتی‌بیوتیکی با روش Kirby-Bauer نسبت به ۱۲ آنتی‌بیوتیک مشخص شد و اطلاعات بدست آمده با نرم‌افزار SPSS مورد بررسی قرار گرفت.

نتایج: تعداد ۶۸ سویه (۳/۲٪) از ایزوله‌ها، آسیتوباکتر بودند. این سویه‌ها بیشتر از بیماران بخش ICU (۲۴ مورد و ۳۵/۸٪) و بخش اورژانس (۱۲ مورد، ۱۷/۹٪) جدا شده بودند و لوله تراشه محل اصلی عفونت بود. کلیستین با ۸۲/۷٪ حساسیت موثرترین آنتی‌بیوتیک بود و بعد از آن افلوکسازین با ۴۷/۴٪ و کلرآمفنیکل با ۳۹/۵٪ حساسیت، قرار داشتند. میزان بالای مقاومت نسبت به مروپنم ۹۸/۱٪ و سفپیم ۹۰/۴٪ مشاهده شد. میزان مرگ و میر ۱۴/۷٪ و بیشتر آن مربوط به سپتی‌سمی بود.

نتیجه‌گیری: با توجه به عفونت‌های جدی و مقاومت بالای آنتی‌بیوتیکی عفونت‌های ناشی از آسیتوباکترها، مراقبت و نظارت مداوم وضعیت حساسیت آنتی‌بیوتیکی ضرورت دارد. همچنین به منظور پیشگیری و کاهش عفونت‌های آسیتوباکتر، التزام جدی به دستورالعمل‌های کنترل عفونت واجب می‌باشد.

کلیدواژه‌ها: آسیتوباکتر، الگوی حساسیت آنتی‌بیوتیکی، عفونت بیمارستانی

نویسنده مسئول:
دکتر صدیقه جوادیپور
مرکز تحقیقات بیماریهای عفونی و
گرمسیری، دانشگاه علوم پزشکی
هرمزگان
بندرعباس - ایران
تلفن: +۹۸ ۷۶ ۲۳۶۶۸۳۵۷
پست الکترونیکی:
sjavadpour@hums.ac.ir

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